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Water quality — Determination of the estrogenic potential of water and waste water —

Part 1: Yeast estrogen screen (*Saccharomyces cerevisiae*)

Qualité de l'eau — Détermination du potentiel oestrogénique de l'eau et des eaux résiduaires —

*Partie 1: Essai d'oestrogénicité sur levures (*Saccharomyces cerevisiae*)*

Reference number
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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

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This document was prepared by Technical Committee ISO/TC 147, *Water quality*, Subcommittee SC 5, *Biological methods*.

A list of all parts in the ISO 19040 series can be found on the ISO website.

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Water quality — Determination of the estrogenic potential of water and waste water —

Part 1:
Yeast estrogen screen (*Saccharomyces cerevisiae*)

WARNING — Persons using this document should be familiar with normal laboratory practice. This document does not purport to address all of the safety problems, if any, associated with its use. It is the responsibility of the user to establish appropriate safety and health practices.

IMPORTANT — It is absolutely essential that tests conducted in accordance with this document be carried out by suitably trained staff.

1 Scope

This document specifies a method for the determination of the estrogenic potential of water and waste water by means of a reporter gene assay with genetically modified yeast strains *Saccharomyces cerevisiae*. This reporter gene assay is based on the activation of the human estrogen receptor alpha.

This method is applicable to:

— fresh water;

— waste water;

— aqueous extracts and leachates;

— eluates of sediments (fresh water);

— pore water;

— aqueous solutions of single substances or of chemical mixtures;

— drinking water.

The limit of quantification (LOQ) of this method for the direct analysis of water samples is between 8 ng/l and 15 ng/l 17β-estradiol equivalents (EEQ) based on the results of the international interlaboratory trial (see Annex F). The upper threshold of the dynamic range for this test is between 120 ng/l and 160 ng/l 17β-estradiol equivalents (EEQ). Samples showing estrogenic potencies above this threshold have to be diluted for a valid quantification. Extraction and pre-concentration of water samples can prove necessary, if their estrogenic potential is below the given LOQ.

2 Normative references

The following documents are referred to in the text in such a way that some or all of their content constitutes requirements of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 3696, *Water for analytical laboratory use — Specification and test methods*

ISO 7027, *Water quality — Determination of turbidity*

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3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

ISO and IEC maintain terminological databases for use in standardization at the following addresses:

— ISO Online browsing platform: available at http://www.iso.org./obp

— IEC Electropedia: available at http://www.electropedia.org/

3.1
blank replicate
additional replicate that contains no test organism, but is treated in the same way as the other replicates of a sample

[SOURCE: ISO 10872:2010, 3.5]

3.2
culture medium
nutrients presented in a form and phase (liquid or solidified) which support microbiological growth

[SOURCE: ISO 6107-6:2004, 24]

3.3
dilution level
D
denominator of the dilution coefficient (using the numerator 1) of a mixture of water or waste water with dilution water as integral number

Note 1 to entry: For undiluted water or waste water, this coefficient per definition is 1→1. The corresponding and smallest possible value of *D* is 1. In this document, the arrow indicates the transition from initial total volume to final total volume.

[SOURCE: ISO 6107-6:2004, 28]

3.4
dilution water
water added to the test sample to prepare a series of defined dilutions

[SOURCE: ISO 20079:2005, 3.7]

3.5
50 % effect concentration
EC_{50}
concentration of a compound which causes 50 % of an effect

Note 1 to entry: In the sense of this document, the EC_{50} is the concentration of a compound which induces 50 % of the maximal reporter gene activity which can be achieved by this compound.

3.6
field blank
container prepared in the laboratory, using reagent water or other blank matrix, and sent with the sampling personnel for exposure to the sampling environment to verify possible contamination during sampling

[SOURCE: ISO 11074:2015, 4.5.3]

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3.7
induction rate
quotient of the mean signal measured after exposure to a dose of the test sample or with a positive control, and the mean signal measured for the negative control using the same experimental conditions

[SOURCE: ISO 6107-6:2004, 43, modified — “corrected absorbance” replaces “mutant colonies”; “wells” replaces “corresponding plates”, “quotient” replaces “difference”.]

3.8
inoculum
fraction of a culture of microorganisms used to start a new culture, or an exponentially growing preculture, in fresh medium

[SOURCE: ISO 6107-6:2004, 44]

3.9
limit of quantification
LOQ
lowest value of a determinant that can be determined with an acceptable level of accuracy and precision

[SOURCE: ISO 15839:2003, 3.18]

3.10
lowest ineffective dilution
LID
lowest dilution within a test batch which does not show any effect, i.e. no statistically significant increase in the reporter gene activity compared with the negative control

[SOURCE: ISO 11350:2012, 3.4, modified — “increase in the reporter gene activity” replaces “increase in the number of revertant wells”.]

3.11
negative control
dilution water without test sample

[SOURCE: ISO 6107-6:2004, 51]

3.12
overnight culture
culture started late in the afternoon and incubated overnight to be ready during the following morning for purposes such as the inoculation of a preculture

Note 1 to entry: The procedure for the overnight culture is described in 9.2.

[SOURCE: ISO 6107-6:2004, 54, modified — deleted: "usually about 16 h".]

3.13
reference compound
compound with one or more property values that are sufficiently reproducible and well established to enable the calibration of the measurement method

[SOURCE: ISO 7405:2008, 3.6, modified — “compound” replaces “material”; “the calibration of the measurement method” replaces “use of the material or substance for the calibration of an apparatus, the assessment of a measurement method or for the assignment of values to materials”.]

3.14
reporter gene activity
quantitative activity of a gene attached to the promoter sequence of another gene

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3.15
stock culture
culture of a strain of organisms maintained under conditions to preserve original features such as nucleotide sequences

[SOURCE: ISO 6107-6:2004, 87]

3.16
test sample
undiluted, diluted or otherwise prepared portion of a sample to be tested, after completion of all preparation steps such as centrifugation, filtration, homogenization, pH adjustment and determination of ionic strength

[SOURCE: ISO 6107-6:2004, 92]

4 Principle

The Yeast Estrogen Screen (YES) is a reporter gene assay which can be used for the measurement of the activation of the human estrogen receptor alpha (hERα) in the presence of a sample containing compounds which activate the estrogen receptor (ER).

By this means the assay detects the estrogenic activity of the whole sample in its actual state as an integral measure including possible additive, synergistic and antagonistic mixture-effects on the whole process of the reporter gene expression.

The basic concept of such assays is explained in References [10] and [11]. The hERα is heterologously expressed in the yeast cell under control of a copper dependent promoter. The estrogen receptor belongs to the family of nuclear hormone receptors. If agonists of the estrogen receptor enter the yeast cell, they bind to the estrogen receptor protein and thus induce its conformational change. As a consequence two receptor proteins form a receptor dimer which translocates to the nucleus. This activation of the estrogen receptor is measured by the induction of the reporter gene *lacZ* which encodes the enzyme β-galactosidase. The *lacZ* is fused to a promoter containing estrogen responsive elements (ERE) and is thus controlled by the activity of the estrogen receptor. The ER-dimer binds to the promoter and by this activates the expression of the β-galactosidase. Finally, the activity of the β-galactosidase as a measure for the estrogenic potential of the sample is determined using an appropriate substrate which is cleaved to a coloured reaction product. The reaction product can be measured photometrically. See Annex C for a scheme of the test principle.

5 Interferences

Coloured or turbid samples might interfere with the photometric detection of cell density and/or the detection of the reaction product of the reporter enzyme β-Galactosidase (see Clause 10 for further information).

Toxic effects of the test sample may lead to a reduction of viable cells and to a reduction of the measurable signal. Consequently, estrogenic effects of a sample may be masked by acute toxic effects leading to false negative test results (see Clause 10 for further information).

High salinity can cause toxic effects due to the resulting osmotic pressure. The conductivity of the sample is a measure for its salinity. The yeast strain constructed by McDonnell et al. (Reference [10]) tolerates a conductivity of the sample up to 34 000 µS/cm.

Bacterial growth in the test wells is assessed by the blank replicate (3.1). See Clause 10 for further information.

If filtered samples are tested in order to remove bacteria from the sample, solid particles are separated from the sample also. Thus, substances with estrogenic activity which are adsorbed on particles might not be detected.

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